8 Answers to Exercises

8.1 Chapter 1 exercises

Workflow

- Identify the NMR active nuclei.
- > Divide these into groups based on chemical equivalence.
- > Everything that follows will be based on these groups of spins.
- > Predict the number of chemical shifts expected in each spectrum.
- Identify the neighbouring groups and predict the splitting each group will cause based on the spins contained in the group.
- > Construct a stick diagram to show the result.
- Q. 1.1 In this exercise we will predict the appearance of some NMR spectra knowing only the spins present. A, B and C have spin ½. X has spin 1 and Y spin ³/₂. Assume that all groups of spins couple to all other groups of spins present.
 - a. Draw stick diagrams to represent the coupling patterns at A in the following spin systems:

Answers

 AC_2 One neighbouring group (C) containing two spin $\frac{1}{2}$ so causes 1:2:1 triplet splitting of

Α

 AB_3 One neighbouring group (B) containing three spin ½ so causes 1:3:3:1 quartet splitting of A

 AB_2X Two neighbouring groups: B contains two spin $\frac{1}{2}$ so causes 1:2:1 triplet splitting of

A; X contains one spin 1 so causes a further 1:1:1 triplet splitting of A.

AB₂**XY** Three neighbouring groups: **B** contains two spin $\frac{1}{2}$ so causes 1:2:1 triplet splitting of **A**; **X** contains one spin 1 so causes a further 1:1:1 triplet splitting of **A**; **Y** contains one spin $\frac{3}{2}$ so causes a further 1:1:1:1 quartet triplet splitting of **A**.

b. What will the coupling pattern at **X** be in **AB**₂**X**? At **Y** in **AB**₂**XY**?

Answers

X in AB₂X Two neighbouring groups: A contains one spin ½ so causes 1:1 doublet splitting of X; B contains two spin ½ so causes a further 1:2:1 triplet splitting of X Y in AB₂XY Three neighbouring groups: A contains one spin ½ so causes 1:1 doublet splitting of Y; B contains two spin ½ so causes a further 1:2:1 triplet splitting of Y; X contains one spin 1 so causes a further 1:1:1 triplet splitting of Y. Q. 1.2 For each of the compounds below:

- a. Identify ALL NMR active nuclei.
- b. If a nucleus is not 100 % abundant, indicate the natural abundance and show the satellites, except for ¹³C, in your answers.
- c. Where quadrupolar nuclei occur, indicate their presence.
- d. Group the nuclei you have identified according to the NMR equivalence and hence decide on the number of chemical shifts that will be found in each nucleus's NMR spectrum.
- e. Draw stick diagrams for the elements specified to indicate where coupling will occur and the splittings that result.

NB In practice the resonances of, and couplings to, quadrupolar nuclei are often not seen. You should ignore quadrupolar nuclei in the rest of this exercise unless you are explicitly asked to comment on them.

NB The NMR spectra of phenyl rings in phosphine ligands are usually complex. You may ignore the phenyl rings in the rest of this exercise. NMR active nuclei are indicated in bold

Answers

^{35/37}Cl are quadrupolar so ignored

Groups are indicated by superscripted labels,

Each group gives rise to a chemical shift

Coupling occurs between groups not within groups

The intensity of a line in a satellite doublet is $\frac{1}{2}$ x natural abundance x the number of equivalent sites in the group causing the satellite (in these examples this is 1) - the intensity is divided between the two lines of the doublet satellite.

Fig. 8. 1 Answers to question 1.2



bis-diphenylphosphinomethane ³¹P 1 chemical shift ¹H 1 chemical shift ¹³C 1 chemical shift, 1%









³¹P 2 chemical shifts

¹³C 2 chemical shifts, 1%







Q. 1.3 Each group of neighbouring coupling spins should be represented by its own layer in the stick diagram; the order in which groups are considered is not important and the horizontal separation of lines in a layer does not need to reflect accurately the size of the coupling constant.









(b)

Fig. 8. 2 Stick diagrams for question 1.3

Answer

The three *groups*, CF_3 , CF_2 , and the second CF_2 should be represented as separate layers in the stick diagrams as in figures (a), (b) and (c)

- a. and (c) correctly show the CF₃ and two CF₂ groups giving a 1:3:3:1 quartet, and two 1:2:1 triplet patterns respectively. The order in which the coupling groups are shown is not important, neither is the magnitude of the "coupling" used, although (a) shows the structure more clearly by choosing separations that avoid overlaps.
- b. incorrectly shows a doublet of triplets of quartets.
- c. does not accurately represent the groups of spins present but shows coupling to four neighbouring groups, CF₃, CF₂, and two CF groups as a result of misrepresenting one of the CF₂ groups as two CF groups having the same coupling constant. Although the splitting pattern that results is "the same" the important and useful information that the triplet arises from a neighbouring group containing two equivalent spin ½ nuclei is obscured.

Although not shown in figure 8.1, it is good practice to indicate the coupling groups responsible for each layer down the side of the stick diagram – see answer to Q1.2 above.



Fig. 8. 3 ¹H NMR spectrum of ¹¹B₂H₆. The complexity is a result not only of coupling to the quadrupolar nucleus ¹¹B but also to the magnetic inequivalence of the terminal protons and borons resulting in a second order pattern

Q. 1.4 Identify which of the following molecules will show second order NMR spectra.



Fluorines are chemically equivalent but magnetically inequivalent, e.g. Fa sees the chemically equivalent protons Ha and Hb differently. Second order

As above, the phosphoruses are chemically equivalent but magnetically inequivalent, they couple differently to the directly bonded and remote

1,2,4,5-tetrafluorbenzene



X = acetylide

 $\begin{matrix} Cl_2 \\ P^A \\ N \end{matrix} \\ \begin{matrix} P^A \\ N \\ II \\ Cl_2 P^A \end{matrix} \\ \begin{matrix} P^A Cl_2 \end{matrix}$

hexachlorotriphosphazene

fac-tris-triphenylphosphinemolybdenumtri(¹³C-carbonyl)



Trans isomer

Second order

195Pt centres Second order

All the fluorines are chemically equivalent and (if we neglect the quadrupolar 14N) do not couple to any other group so are magnetically equivalent. Compare with the proton NMR spectrum of ethene. First order

Gauche isomer

FA and FA' are chemically equivalent but couple differently to FB and FB'. FA has a geminal coupling to FB 2J(FAFB) but a vicinal coupling to FB', 3J(FAFB'). FA and FA' are magnetically inequivalent Second order

 $\overset{H^{t''}}{\overset{}_{H^{t'''}}} \stackrel{B^{A'}}{\overset{}_{H^{b''}}} \overset{H^{b''}}{\overset{}_{H^{b''}}} \overset{B^{A'}}{\overset{}_{H^{t''}}} \overset{H^{t''}}{\overset{}_{H^{t'''}}}$

tetrafluorohydrazine

diborane

Borons are chemically equivalent, Ht are chemically equivalent, Hb are chemically equivalent. Consider Ht. This has a one bond coupling to BA' but a three bond coupling to BA; Ht to Ht''' are magnetically inequivalent as are BA and BA'. Second order

Coupling to quadrupolar chlorine and 14N can be ignored Phosphoruses are chemically and magnetically equivalent First order

Each phosphorus "sees" one 13CO trans but the other two cis. Phosphoruses are chemically equivalent but magnetically inequivalent, 13CO are chemically equivalent but magnetically inequivalent

Phosphoruses are chemically equivalent 13CO ligands are chemically equivalent

<Unnumbered Chemdraw 8.1>

Q. 1.5 Write down a stick diagram that represents the couplings present in the ¹H NMR spectrum of [Cp₂W(H)(SnCl(t-Bu)₂)], hydride region only. Exclude coupling to the Cp rings.

Answer

`w ^{, , , н} Sn(t-Bu)₂Cl

The hydride ligand will couple to tin and tungsten, figure 8.4.

<Unnumbered Chemdraw 8.2>

¹⁸³W is spin ½, 14.4% abundant so will give satellite peaks, a doublet of total intensity 14.4
%, i.e. each peak in the doublet will have intensity 7.2%.

Tin has two isotopes of sufficient abundance to give clearly visible satellites ¹¹⁷Sn (7.6%) and ¹¹⁹Sn (8.6%), there will be two sets of tin satellites. Both ¹¹⁷Sn and ¹¹⁹Sn are spin ½ so both sets of satellites will be doublets.

The lines in the ¹¹⁷Sn satellites will have intensity 7.6/2 = 3.8% and in the ¹¹⁹Sn satellites 4.3%. The unsplit central line will have intensity 100-14.4-7.6-8.6 = 69.4%.

Isotopologues containing NMR active tin and tungsten will give satellites on the satellites of very low intensity, e.g. $(14.4 \times 8.6)/2 = 0.6\%$. These can just be seen in the

experimental spectrum and are indicated by a stick diagram below the experimental spectrum.



Fig. 8. 4 Assignment of the ¹H NMR spectrum of [Cp₂W(H)(SnCl(t-Bu)₂)]. Experimental spectrum reproduced from Mobley, T. A., *et al., Magn. Reson. Chem.*, 2010, 48, 787–792. doi:10.1002/mrc.2663, Copyright © John Wiley & Sons.

8.2 Chapter 2 exercises

Q. 2.1 The ³¹P{¹H} NMR spectra of the dimeric phosphinidene {M(μ-PAr)}₂ 1, 2, 3 and the ²⁰⁷Pb{¹H} spectrum of 2 are shown in figure 2.9. Account for the all couplings observed. Suggest a reason for the inability to observe the ⁷³Ge NMR spectrum.

Answers

 ${Ge(\mu-PAr)}_2$ One symmetrical pattern is seen in the ³¹P{¹H} NMR spectrum, i.e. one phosphorus chemical shift so one type of P is present. The resonance is a singlet with satellites due to coupling to ⁷³Ge, abundance 7.76%, I = 9/2. The satellites are a 1:1:1: etc decet indicating each P couples to only one ⁷³Ge. The intensity of each line in the decet is 7.76/10 = 0.776% of the total intensity, or ca. 0.84% of the central line. Coupling to only one ⁷³Ge is expected, the chances of both germanium atoms in the molecule being ⁷³Ge is 0.0776x0.0776 or 1 in six thousand. These resonances will be too weak to be seen in the spectrum.

The ⁷³Ge spectrum cannot be observed due to the large quadrupole moment of ⁷³Ge and low natural abundance. ⁷³Ge NMR resonances are expected to be very low intensity compounded by being very broad due to efficient relaxation causing large Heisenberg broadening at the low symmetry Ge site.

(Sn(\mu-PAr))₂ One symmetrical pattern is seen in the ³¹P{¹H} NMR spectrum, i.e. one phosphorus chemical shift so one type of P is present. The resonance is a singlet with satellites due to coupling to ¹¹⁷Sn and ¹¹⁹Sn, abundance 7.68% and 8.59%, I = $\frac{1}{2}$. Satellites due to ¹¹⁵Sn are too weak to be observed The satellites are a 1:1 doublets indicating each P couples to only one ^{11x}Sn. The intensity of each line in the doublets is ca. 8/2 = 4% of the total intensity, or ca. 5% of the central line. Coupling to only one ^{11x}Sn is expected, the chances of both tin atoms in the molecule being ^{11x}Sn is ca. 0.0768x0.0859 or around 1 in six thousand. These resonances will be too weak to be seen in the spectrum.

One symmetrical pattern is seen in the ¹¹⁹Sn{¹H} NMR spectrum, i.e. one tin chemical shift so one type of Sn is present. The resonance is a triplet indicating the tin couples to two equivalent phosphorus centres. Satellites due to coupling to ¹¹⁷Sn are not seen due to the low abundance of the isotopologue containing ¹¹⁹Sn at one site and ¹¹⁷Sn at the other. If both tin nuclei are ¹¹⁹Sn then the tins are equivalent so do not couple.

{**Pb**(μ -**PAr**)}₂ One symmetrical pattern is seen in the ³¹P{¹H} NMR spectrum, i.e. one phosphorus chemical shift so one type of P is present. The resonance is a singlet with two sets of satellites due to coupling to ²⁰⁷Pb, abundance 22.1%, I = 1/2. The more intense satellites are a 1:1 doublet indicating each P couples to only one ²⁰⁷Pb. The intensity of each line in the doublet is ca. 22/2 = 11% of the total intensity, or ca. 14% of the central line. The weaker satellites are due to the isotopologue in which both leads are ²⁰⁷Pb, abundance ca. 0.22x0.22 or around 5 in a hundred. The satellites due to this isotopologue will be a triplet (2 equivalent I = $\frac{1}{2}$ neighbours). The outer lines of the triplet can just be seen, the central line is underneath the main peak in the spectrum. One symmetrical pattern is seen in the 207 Pb{ 1 H} NMR spectrum, i.e. one lead chemical shift so one type of lead is present. The resonance is a triplet indicating the lead couples to two equivalent phosphorus centres. If both leads are 207 Pb the two leads are equivalent so do not couple. No satellite peaks are present.

Q. 2.2 Reaction of [PtH(dtbpx)(MeOH][OTf]₂ A with ¹³CO affords B. The NMR data for A and B are given in Table 2.1. The ¹H and ³¹P{¹H} NMR spectra are shown in Figs. 8.6 and 8.7.

	δ /ppm ^a	<i>J</i> (P¹-	<i>J</i> (P²-	J(Pt-⁵			
Compound							
Α							
P ¹	46.2 d	-	6	4312			
P ²	34.6 d	6	-	2095			
Н	-7.3 dd	18	176	805			
Compound							
В							
P ¹	42.2 dd	-	19	2958			
P ²	33.2 dd	19	-	2011			
Hc	-4.2 ddd	15	145	740			
СО	179.3 dd	113	8	1299			

Table 8.1 Selected NMR data for the complexes

^a All spectra recorded with ¹H decoupling, where given couplings to ¹H were determined from the ¹H NMR spectrum. ^{b 195}Pt satellites are seen on all resonances. ^c J(HC) not determined. d = doublet.

- a. Use the fact that first order coupled multiplets are symmetrical to assign the ¹⁹⁵Pt satellites to their corresponding central peaks in the ³¹P{¹H} NMR spectra in figure 2.10
- b. Use the magnitudes of the various coupling constants to determine the stereochemistry around the Pt centre.
- c. Suggest a reason for the roofing observed in the ${}^{31}P{}^{1}H$ NMR spectrum of **B**.

Answers, figures 8.5 and 8.6

Trans couplings are larger than *cis* couplings. The H is *cis* to P¹ and *trans* to P² in both A and B. The CO is *trans* to P¹ and *cis* to P² in B.

Roofing indicates the onset of second order effects and is seen when two coupled nuclei have very similar chemical shifts, the difference in shifts being comparable to the coupling constant.

Coupling to ¹H is not seen in the ${}^{13}C{}^{1}H$ and ${}^{31}P{}^{1}H$ NMR spectra due to proton decoupling.



Fig. 8. 5 ¹H NMR spectra of compounds **A**, and **B** showing the assignment of the ¹⁹⁵Pt satellites. Solid and dotted lines are used to indicate two branches of the same set of satellites.



Fig. 8. 6 ³¹P{sel-¹H} NMR spectra of compounds A, and B showing the assignment of the ¹⁹⁵Pt satellites. Solid and dotted lines are used to indicate two branches of the same set of satellites. Selective proton decoupling has been used to remove couplings to phenyl protons. The aromatic region of the ¹H NMR spectra are not shown. A slight second order distortion can be seen in the upfield satellites of the ¹H and ³¹P{¹H} NMR spectra of B. The latter also contains accidental overlapping of lines the upfield satellites of the two different phosphoruses in B.

Q. 2.3 Advanced question.

Interpret fully the spectra in Figs. Answers Q2.1&2 and Table 2.1 and hence confirm the identities of **A** and **B**.



< Unnumbered Chemdraw 8.3>

Answer

Compound A

¹H NMR spectrum. One resonance is seen in the metal hydride region, i.e. one hydride chemical shift, so one type of hydride ligand is present. The splitting pattern is a doublet of doublets with doublet satellites due to coupling to ¹⁹⁵Pt, so the hydride has two neighbouring groups in addition to platinum. The doublet splittings assigned to *J*(PH) in the table tells us there is one P¹ and one P² in the compound while the doublet splitting of the satellites tells us there is one Pt. The coupling of 176 Hz assigned to *J*(P²H) is much larger than that assigned to *J*(P¹H) (18 Hz) so P² is *trans* to the hydride and P¹ is *cis*, *J*_{trans} >> *J*_{cis}.

³¹P{¹H} NMR spectrum. Two resonances are seen, i.e. two phosphorus chemical shifts, so two types of phosphorus are present. The resonances are doublets with satellites due to coupling to ¹⁹⁵Pt, so each has one neighbouring group in addition to platinum. A coupling of 6 Hz appears in both doublets so is $J(P^1P^2)$. The doublet splitting tells us there is one P¹ and one P² in the compound. We are told the ¹⁹⁵Pt satellites are doublets, so one Pt is present confirming our deductions from the ¹H NMR spectrum. ¹ $J(P^1Pt) >>$ ¹ $J(P^2Pt)$ indicating a low *trans* influence ligand (MeOH) is *trans* to P¹ and a high *trans* influence ligand (hydride) is *trans* to P² consistent with our assignment from the proton NMR spectrum.

Chemical common sense tells us that there can only be one hydride present in the complex; Pt(II), d⁸ ion so we expect a square planar complex. The ligands *trans* to each phosphorus must be different since the Ps are different, so cannot both be H. We are given the molecular formula of **A** which confirms our assignment of the spectra. Analysis of the coupling constants allows unequivocal assignment of the stereochemistry.

Compound B

¹³C{¹H} NMR spectrum. One resonance is seen in the metal carbonyl region, i.e. one carbon chemical shifts, so one type of carbonyl ligand is present. The splitting pattern is a doublet of doublets with doublet satellites due to coupling to ¹⁹⁵Pt, so the CO has two neighbouring groups, each containing one spin ½ nucleus in addition to the group containing one platinum. Matching the coupling constants tells us these are P¹ and P². The doublet splittings tells us there is one P¹, one P² and one Pt in the compound. The

coupling of 113 Hz is assigned to $J(P^1C)$ and of 8 Hz to $J(P^2C)$ so P¹ is *trans* to the CO and P² is *cis* – $J_{trans} >> J_{cis}$.

³¹P{¹H} NMR spectrum. Proceeding as above can assign the coupling of 19 Hz that appears in both multiplets to $J(P^1P^2)$ and deduce that there is one P¹ and one P² in the complex. We note that an extra doublet coupling is seen on all resonances indicating that one ¹³CO ligand is present in the complex in addition to the two inequivalent phosphorus ligands.

Both ¹*J*(PtP) are small indicating both Ps are *trans* to high *trans* influence ligands. ¹*J*(PtP²) in **A** and **B** are comparable suggesting P² is *trans* to the hydride in both complexes.

¹**H NMR spectrum.** Proceeding as above we can deduce that there is one type of hydride present coupling to two phosphorus groups each containing one phosphorus and to one platinum. We can guess the third doublet splitting arises from coupling to ¹³CO and we are told *J*(CH) was not determined. The large *J*(PH) coupling can be assigned to P², the phosphorus *trans* to H while the smaller *J*(PH) coupling can be assigned to the *cis* P¹. This assignment is consistent with our assignment of the *J*(PPt) couplings above.

Q. 2.4 Advanced question.

The three spectra shown in figure 8.8 each show a symmetrical six line pattern in the upfield region yet each arises from a different spin system.

The first spin system comprises: group A containing two equivalent spins and two groups, B and C, containing one spin $\frac{1}{2}$ nuclei. B and C have similar chemical shifts and do not couple to each other but do couple to group A with, accidentally, the same coupling constant J(AB) = J(AC). B and C are of the same element but A is a hetero nucleus.

The second spin system comprises: group A containing one spin ½ nucleus and two groups, B and C, also containing one spin ½ nuclei. B and C have similar chemical shifts and couple to each other and to group A, all the coupling constants are, accidentally, the same. A is a hetero nucleus.

Finally, the third spin system comprises: groups A and B each containing one spin ¹/₂ nucleus and group C containing two spin ¹/₂ nuclei. B and C have dissimilar chemical shifts and couple to each other. B also couples to group A but C does not. All the coupling constants are different. B and C are of the same element but A is a hetero nucleus.

Using the workflow in Chapter 2 and paying particular attention to the caveats and information in the side bar box, explain which spectrum corresponds to which spin system and how these can be differentiated.

Answers

Spectrum (a) contains two symmetrical patterns indicating two chemical shifts are present, so two groups of spins are expected. However, the upfield resonance is a doublet of triplets indicating coupling to two groups of spins implying three groups of spins must be present in the molecule. The third group does not appear in the spectrum, so must be a heteronuclear group of spins.

The doublet coupling in the downfield multiplet matches the triplet coupling in the upfield multiplet, so the downfield group must contain two spin $\frac{1}{2}$ nuclei and the upfield group one spin $\frac{1}{2}$ nucleus.

The large doublet coupling must then be the heteronuclear coupling, so the heteronuclear group must contain one spin ½. Spectrum (c) corresponds to the last spin system.

Spectrum (b) contains a single pattern showing distinct roofing. Roofing is commonly seen when two groups of spins having similar chemical shifts couple to each other. This corresponds to groups B and C of the second spin system in the question. Coupling to the hetero nuclear spin A splits the AB quartet. The accidental coincidence of all the coupling constants gives the appearance of a roofed doublet of triplets.

Spectrum (c) shows a single, undistorted apparent doublet of triplets. This would imply two heteronuclear neighbouring groups, one containing two spin $\frac{1}{2}$ nuclei the other one spin $\frac{1}{2}$. None of the spin systems correspond to this.

However, we note that the first spin system contains two groups A and B that have similar chemical shifts but do not couple. In the absence of any other groups of spins, this would

give two singlets. If we introduce a third group, A, which is heteronuclear, containing two spin $\frac{1}{2}$ nuclei, the singlets would each split into triplets. Since J(AB) = J(AC), an apparent doublet of triplets results.



We thank a reviewer for suggesting these situations.

Fig. 8. 7 Simulated NMR spectra for Q2.4.

8.3 Chapter 3 exercises

- **Q 3.1** Suggest two ways in which NMR measurements might be used to determine order the basicity of a library of phosphine ligands.
 - a. The magnitude of the selenium-phosphorus coupling in organophosphorus selenides, R₃PSe, increases as the basicity of the phosphine, decreases, Me < Ph < OMe, Figure 3.15.
 - b. As the phosphine becomes more basic/electron donating from P(OPh)₃ to PBu₃, the ¹³CO chemical shift increases; see figure 3.6 which plots the ¹³CO chemical shift against the Tolman electronic parameter of the phosphorus ligand or CO stretching frequency for two series of complexes W(CO)₅L and CpFe(CO)(PR₃)(SnPh₃).
 - c. ¹J(PtP) in cis-PtCl2(PR₃)₂ reflects the basicity of the phosphorus ligand being 3508 Hz in cis-PtCl₂(PBu₃)₂ and cis-PtCl₂{P(OEt)₃}₂ is 5698 Hz. Interestingly, the ratio of the phosphine and phosphite coupling constants is fairly independent of the nature of the acceptor; being 1.62 in these platinum complexes and 1.53 in the borane complexes Me₃P-BH₃, ¹J(¹¹BP) 64 Hz, and (MeO)₃P-BH₃, 97 Hz
- **Q 3.2** What is meant by "paramagnetic term" in discussing the chemical shift of transition metals?

Answer The term refers to shielding caused by mixing in of excited states, this can cause either shielding or deshielding.

Q 3.3 A series of Pt complexes **1** has been prepared where one of X and Y is a methyl group and the other is one of the following ligands: acetone, acyl, SnCl₂ or CO. ${}^{1}J({}^{195}Pt^{31}P)$ in the complexes varies from 3367 to 3995 to 4036 to 5003. By considering the effect of *trans*-influence, decide if X or Y is the methyl group. Pair each ligand with the appropriate ${}^{1}J({}^{195}Pt^{31}P)$.

Answer ${}^{1}J({}^{195}Pt^{31}P)$ is strongly influenced by the *trans* ligand. Since ${}^{1}J({}^{195}Pt^{31}P)$ is observed to vary widely between the complexes, it is reasonable to assume that it is the ligand *trans* to the phosphine that is varying, hence Y is the invariant Me ligand. The order of *trans*-influence in the complexes is CO > acyl > SnCl₂ > acetone which is the order in which ${}^{1}J({}^{195}Pt^{31}P)$ increases.

<Unnumbered Chemdraw 8.4>



Q 3.4 Sketch stick diagrams to illustrate the couplings present in the ¹H NMR spectrum of each isotopologue of **2**. ¹*J*(¹⁰⁹Ag¹H) in the dinuclear silver compound **2** is 134 Hz. Use the data in Table 1.2 to calculate ¹*J*(¹⁰⁷Ag¹H). From data in B. K. Tate *et al.*, Chem. Sci. 2013, **4**, 3068.

Answer For coupling over bonds in which only the isotopes of the coupling partners have changed, the value of *J* scales as the gyromagnetic ratios. γ ⁽¹⁰⁷Ag) = -1.088918 x 10⁷ rad.s⁻¹.T⁻¹ and γ ⁽¹⁰⁹Ag) = -1.2519 x 10⁷ rad.s⁻¹.T⁻¹ so ¹*J*(¹⁰⁷Ag¹H) =116.6 Hz.

Both isotopes of silver are spin $\frac{1}{2}$ so in the ¹⁰⁷Ag¹⁰⁷Ag and ¹⁰⁹Ag¹⁰⁹Ag isotopologues the proton will split into a triplet (two equivalent spin $\frac{1}{2}$ neighbours) with ¹J(¹⁰⁷Ag¹H) 116.6 Hz and ¹J(¹⁰⁹Ag¹H) 134 Hz respectively. The silvers are chemically equivalent, so we cannot distinguish the ¹⁰⁷Ag¹⁰⁹Ag isotopologue from the ¹⁰⁹Ag¹⁰⁷Ag isotopologue the proton is split into a doublet of doublets ¹J(¹⁰⁷Ag¹H) 116.6 Hz and ¹J(¹⁰⁹Ag¹H) 134 Hz. It does not matter which coupling is taken first in the stick diagram. The isotopic abundance of ¹⁰⁷Ag = 51.8% and of ¹⁰⁹Ag = 48.1%, which for simplicity we can take as 50:50.



Fig. 8.8 Stick diagrams for Q3.4

8.4 Chapter 4 exercises

Q 4.1 There is one, isolated group of 19F spins in CF₃Br. Before the 90° pulse, the magnetization will be aligned along the z-axis in the rotating frame. The -x pulse rotates the magnetization through 90° in a clockwise direction (looking back along x to the origin) along the x-axis. The magnetization now lies in the xy plane and is aligned along the y-axis. After the pulse the magnetization will start to precess in the xy plane at a rate that depends on the chemical shift offset from the Larmor frequency. Since there is only one chemical shift and no *J* coupling a single, precessing magnetization vector is seen, Fig. 8.9.



Fig. 8.9 Effect of a $\pi/2$ x pulse and subsequent evolution of the magnetization of an isolated group of spins.

Q 4.2 In CHF₃ coupling between the H and F, splits the ¹⁹F resonance into a doublet. There will be two magnetization vectors that will precess at different rates, +/-J(HF)/2, i.e. separated by J(HF), Fig. 8.10.



Fig. 8.10 Effect of a $\pi/2$ -x pulse and subsequent evolution of the magnetization of a group of spins with one spin $\frac{1}{2}$ neighbour. The effect of chemical shift (Fig. 4.7) is omitted for clarity.

Q 4.3 The 90° -y-pulse rotates the magnetization to the -x-axis. (NB It's easier to rotate the cartesian axes in the figure, making the y-axis come out of the paper towards us than to draw the magnetization vectors behind the plane of the paper.) After time τ the two branches of the doublet, shown as a solid and a dotted arrow, will have split and lie between -x and y and -x and -y. For clarity we now look down along z-axis, the xy plane lies in the paper. The 180° x-pulse flips the magnetization vectors about the x-axis so that the vector that was between -x and y is now between -x and -y and *vice versa*. However, the direction in which the two vectors precess does not change, so after a further time τ the vectors come back together along -x, figure 8.11



Fig. 8. 11 The effect of the pulse sequence is to form a spin echo. The magnetization refocuses along the -x axis. The effect of chemical shift (Fig. 4.7) is omitted for clarity.

Q 4.4 The "zero-crossing" times, t, occurs when the magnetization has relaxed back along the z-axis to the origin, i.e. at the "halfway" point. For a first order process, rearranging Eq. 4.3 and solving for $M_t = 0$ shows this occurs when t = T₁ln2.

PPr₃	PBu₃	P(C ₆ H ₁₁) ₃	P(Me ₂ Ph) ₃	P(MePh ₂) ₃	P(<i>p</i> -tol)₃
4.4	3	2.5	9.6	7.5	7.2
6.4	4.3	3.6	13.9	10.8	10.4
	PPr ₃ 4.4 6.4	PPr3 PBu3 4.4 3 6.4 4.3	PPr3 PBu3 P(C6H11)3 4.4 3 2.5 6.4 4.3 3.6	PPr3 PBu3 P(C6H11)3 P(Me2Ph)3 4.4 3 2.5 9.6 6.4 4.3 3.6 13.9	PPr3 PBu3 P(C6H11)3 P(Me2Ph)3 P(MePh2)3 4.4 3 2.5 9.6 7.5 6.4 4.3 3.6 13.9 10.8

The faster spin-lattice relaxation observed for the trialkyl phosphines is attributed to dipole-dipole relaxation of the ³¹P spins by the adjacent proton spins, which are more numerous in the trialkyl phosphines than in the aryl phosphines.

8.5 Chapter 5 exercises

- Q 5.1 Figure 5.20 shows the structure and the ¹¹⁹Sn{¹H} refocused INEPT NMR spectrum of the polyoxometalate anion [(MeO)SnW₅O₁₈]³⁻ in which there are two tungsten environments, W_{eq} and W_{ax}. The nuclear spin and abundance of the isotopes of tin and tungsten are given in Table 1.2.
 - Explain why the INEPT experiment is preferred rather than a simple proton decoupling experiment for observation of ¹¹⁹Sn?
 - Assign the couplings seen in the ¹¹⁹Sn{¹H} spectrum and account for the observation that the main satellite sub-spectrum is a doublet.
 - c. Predict the appearance of the $^{183}W{^1H}$ spectrum.

Answer

a. The INEPT experiment affords a signal enhancement of γH/γSn ≈ 2.7. Further signal:noise enhancement is available since the scan repetition rate is determined by the faster, proton relaxation allowing more scans to be acquired in the same experiment time.

b. The natural abundance of ¹⁸³W is 14.31%. The chances of having two equatorial ¹⁸³W_{ax} are thus 0.14x0.14x6 (number of different ways of placing two ¹⁸³W_{ax} in equatorial sites and ignoring double counting from 3 or $4x^{183}W_{ax}$ isotopologues) in the molecule are ~12%. This intensity would be split into a 1:2:1 triplet centred on the "no ¹⁸³W_{ax}" peak at δ = -647 ppm. The central peak of the triplet lies beneath the "no ¹⁸³W_{ax}" peak and the outlying peaks have 3% of the total spectral intensity and are at located at ¹*J*(¹¹⁹Sn¹⁸³W_{eq}) from the central line and can be seen at ca. -646.8 and -647.2 ppm), figure 8.12.



Fig. 8.12. Assignment of ¹¹⁹Sn{¹H} refocused INEPT NMR spectrum of [(MeO)SnW₅O₁₈]³⁻.
Adapted with permission from B. Kandasamy *et al.*, *Chem. Eur. J.*, 2012, **18**, 59–62.
© 2011 WILEY-VCH Verlag GmbH & Co.

c. The ¹⁸³W{¹H} NMR spectrum will show two chemical shifts δ_{ax} and δ_{eq} in the ratio 1:4. Both will show satellites due to coupling to ¹¹⁵Sn, ¹¹⁷Sn, and ¹¹⁹Sn. Each set of satellites will be a doublet. The satellite sub-spectra will have intensities 0.17%, 3.84% and 4.29% respectively, i.e. half the natural abundance of each isotope of tin since the total intensity in each sub-spectrum is split in two on forming a doublet. The close similarity of χ (¹¹⁹Sn) and χ (¹¹⁷Sn) results in the doublets due to ¹J(¹¹⁹Sn¹⁸³W) (38 Hz) and ¹J(¹¹⁷Sn¹⁸³W) (36 Hz) being unresolved due to the linewidth (> 2 Hz).

Q 5.2Figure 8.13 shows the ³¹P,³¹P-COSY of (NacNac)Rh(P₅Ph₅).

- a. Identify the number of distinct phosphorus sites in the molecule.
- b. Using the information about coordination shifts given in chapter 2, identify the resonances of P2 and P3.
- c. Hence assign the remaining correlations in the spectrum and attempt an assignment of the couplings in the 1D $^{31}P{^{1}H}$ spectrum along on the F1 and F2 axes.



Fig. 8.13. ³¹P,³¹P-COSY for (NacNac)Rh(P₅Ph₅). Correlations between A and E (dotted line), A and D (dashed line) and ADCB (solid line) are indicated. Adapted from S. J. Geier and D. W. Stephan, *Chem. Commun.*, 2008, 2779–2781 with permission from The Royal Society of Chemistry.

- a. 5³¹P shifts are seen so 5 distinct types of phosphorus are present, figure 8.13.
- b. Coordination to a transition metal is expected to result in a down field shift of the phosphorus resonances suggesting P3 and P2 give the resonances at ca. 50 (A) and 30 (B) ppm. P2 and P3 will also show ¹J(RhP) whereas P1, P4 an P5 will show smaller ²J(RhP) confirming the assignment. The apparent triplet substructure in the multiplets A and B is due to the accidental degeneracy of two coupling constants in each multiplet.
- c. Resonance E couples to both A and B suggesting E is P1. A also couples to D which couples to C which couples to B indicating ABCD are either P2P4P5P3 or P3P5P4P2, the data does not distinguish these possibilities. Weak correlations attributed to long range couplings are also seen between D and E and C and E.
- **Q 5.3** Figure 8.14 shows the ¹H,¹³C-HMQC NMR spectrum of the 1,1-dimethyl-1,5-dithiol-3-thiapentane oxorhenium complex shown. The ¹³C spectrum has been fully assigned.
 - a. Account for the occurrence of eight sets of resonances in the ¹H NMR spectrum.
 - b. Given only ${}^{1}J_{CH}$ correlations are observed, assign the ${}^{1}H$ spectrum indicating to which carbon in the backbone each proton is bonded.
 - c. How might C(1) and C(2) be distinguished?

Answer

- a. The proton/methyl group on each carbon is directed either beneath the S₃I plane
 i.e. under the Re centre (*exo*, indicated with primes), or towards the O atom
 (*endo*). The *endo* protons are deshielded by the Re=O bond.
- b. See figure. Only the exo- and endo-Me groups are shown for clarity
- c. C(1) and C(2) can be distinguished by an HMBC spectrum in which a long range correlation is seen between C(3) and H(2') but not between C(3) and H(1) or H(1').



Fig. 8.14. (a) ¹H,¹³C-HMQC NMR spectrum of chiral oxorhenium complex, containing 1,1-dimethyl-1,5-dithiol-3-thiapentane ligands in acetone-*d*6. A residual acetone peak (*) can be seen at 2.05 ppm. (b) Fragment of HMBC spectrum, correlation between C(2) and H(3') indicated. Doublets are due to ¹*J*(CH) correlations. Adapted from F. De Montigny *et al.*, *Phys. Chem. Chem. Phys.*, 2010, **12**, 8792–8803 with permission from the PCCP Owner Societies.

- Q 5.4 Figure 8.15 shows part of the NOESY spectrum of the reaction product from the addition of ¹³C-enriched MeReO₃ to [(bpy)PtMe₂] (bpy = 2,2'-bipyridine) recorded at ¹H = 500 MHz.
 - a. Identify the resonances of the $^{13}CH_3$ group.
 - b. Use the NOESY correlations to assign the correlations observed.
 - c. Hence decide if *cis* or *trans* oxidative addition of MeReO₃ to [(bpy)PtMe₂] has occurred.

- a. The spectral lines, labelled F, at ca 0.05 and 0.31 ppm have half the intensity of the other lines and are separated by ca 130 Hz, both observations are consistent with these lines being the ¹³C coupled doublet of the ¹³CH₃ protons, figure 8.15.
- b. The resonances F show a NOESY correlation (short dashes) to the resonance E at 1.4 ppm but not to the resonance D at 0.18 ppm placing the ¹³CH₃ group adjacent to E but remote from D. D shows a NOESY correlation (long dashes) to E indicating D is adjacent to E. Finally, the chemical shift of D is similar to that of the ¹³CH₃ doublet F suggesting both methyl groups occupy similar environments at Pt. These data are consistent with F and D being remote from each other, i.e. occupying axial sites, while E is *cis* to both, i.e. occupies an equatorial site.





Fig. 8.15. NOESY spectrum of the reaction product from addition of ¹³C-enriched MeReO₃ to [(bpy)PtMe₂] (bpy = 2,2'-bipyridine). Adapted with permission from K. H. Pichaandi *et al.*, *Inorg. Chem.* 2017, **56**, 2145–2152. Copyright © 2017, American Chemical Society.

8.6 Chapter 6 exercises

Q 6.1. Q1. Figure 6.19 shows the variable temperature ³¹P NMR spectra of $[Ni(L)_2]$, L = SeSPPh₂. Suggest a dynamic process that accounts for the high and low temperature spectra.

Answer Two isomers exist of the square planar complex. In one the S donors are mutually *cis* in the other they are mutually *trans*. At high temperature, rapid interchange of the isomers occurs *via* a tetrahedral intermediate, while at low temperature interchange is frozen out. Note the ⁷⁷Se satellites visible at low and high temperature.

Q 6.2. Figure 6-20 shows the ¹⁹F NMR spectra of AgBF₄ in acetone recorded at low, intermediate and high temperature. Explain why two chemical shifts, with different coupling patterns and intensities, are expected at low temperature. Suggest an explanation for the collapse of the resonances to a singlet, at a shift closer to low T quartet than to the low septet at high T. Suggest two possible reasons why the coupling in the septet is lost before that in the quartet.

Answer Boron has two NMR active isotopes, ¹⁰B (~20%, spin 3) and ¹¹B (~80%, spin $\frac{3}{2}$). At low temperature separate resonances are seen for [¹⁰BF₄]⁻ and [¹¹BF₄]⁻ at slightly different chemical shift due to zero point energy effects – an isotopologue shift. ¹⁰B splits the ¹⁹F into seven equally intense lines (2*I*+1) while ¹¹B splits the fluorine resonance into four lines. As the temperature is raised, exchange of fluorine between [¹⁰BF₄]⁻ and [¹¹BF₄]⁻ occurs and an exchange decoupled resonance at the weighted average chemical shift is seen.

The magnitude of a coupling constant scales with the gyromagnetic ratio for isotopologues of same coupling partners; $\gamma^{11}B = 8.58406$, $\gamma^{10}B = 2.67085 \ 10^7 \text{radT}^{-1}\text{s}^{-1}$ so $J(^{11}B^{19}\text{F})$ is ~ 4 times $J(^{10}B^{19}\text{F})$.

As the bond between B and F breaks, exchange decoupling occurs. Since $[^{10}BF_4]^-$ is less abundant than $[^{11}BF_4]^-$ its rate of site leaving must be higher. Furthermore, $J(^{10}B^{19}F)$ is smaller than $J(^{10}B^{19}F)$, so requires a lower rate of site leaving to be exchange decoupled. These factors work together and coupling to ^{10}B is lost at a lower rate of exchange (lower T) than coupling to ^{11}B .

Q 6.3. Account fully for the temperature dependence of the ¹³C NMR spectra of [Rh(PBu₃)₂(CO)CI] in the presence of traces of dissolved CO, figure 6.21.

Answer In the absence of CO the ¹³C NMR spectrum of [Rh(PBu₃)₂(¹³CO)CI] shows a well resolved doublet of triplets due to coupling to the two equivalent phosphines and ¹⁰³Rh. Addition of traces of CO results in loss of the small coupling to phosphorus, simplistically, the rate of bond breaking is greater than the phosphorus-carbon coupling constant so the coupling disappears, the phosphorus is exchange decoupled from the carbon. This is

consistent with the on-set of *intermolecular* exchange of free and co-ordinated CO giving exchange decoupling and loss of ${}^{2}J_{PC}$.

The larger rhodium-carbon coupling, however, is still present, even though it is the rhodium-carbon bond that is breaking, since the rate of bond cleavage does not yet exceed this coupling constant. At high CO concentration, exchange of free and coordinated CO becomes fast enough to decouple the rhodium and carbon nuclei and a singlet is seen.

Q 6.4.1 Figure 6.22-left shows the variable temperature ${}^{31}P{}^{1}H{}NMR$ spectrum of a mixture of [PPh₃Cl]Br and [PPh₃Cl]Br. Account for the observation of separate resonances at -60 °C but a single resonance at +20 °C.

Answer The ³¹P{¹H} VT NMR shows interconversion of [PPh₃-Cl] Br and [PPh₃-Br] Cl is slow, (low temperature limit) at -60 °C. As T is raised, the rate of interconversion increases until coalescence is reached at 0 °C. Above 10 °C exchange is fast, a single average resonance is seen which narrows as the exchange rate increases.

Q 6.4.2 Figure 6.22-centre also shows the EXSY spectrum of a mixture **1**-X. Identify which species participate in exchange and assign the exchanging pairs. By considering the chiral information decide if exchange occurs with retention or inversion at phosphorus.

Answer The peaks at 68/72 and 72/68 ppm correspond to bromide self-exchange. The four remaining peaks correspond to cross-exchange between bromo and chloro species. *S*-1-Cl is paired with *R*-1-Br and *S*-1-Br with *R*-1-Cl. No cross-peaks are seen linking *S*-1-Cl is paired with *S*-1-Br or *R*-1-Br with *R*-1-Cl; nucleophilic exchange in halophosphonium species occurs with inversion of configuration at P.

8.7 Chapter 7 exercises

Q 7.1 Figure 8.16 shows the ¹⁰³Rh{¹H} CP/MAS NMR spectrum of the symmetrical dimeric Rh cation [$(H_2O)_4$ Rh(μ^2 -OH)₂Rh($H_2O)_4$], recorded at two different MAS rates (1.0 kHz, top; 4.0 kHz.

- a. At what frequency, v_0 , were the spectra recorded?
- b. Can the CSA principal values of δ_{11} , δ_{22} , δ_{33} , and δ_{1so} be estimated from the spectra? If so, give approximate values.
- c. Explain why the intensity of the CP spectrum first increases then dies away.

Answer

a. For this sample 4 kHz is a high enough spinning rate for a resonance at the isotropic chemical shift to be observed. The remaining side bands occur on either side of the isotropic peak offset at a frequency equal to the spinning rate, 4 kHz.

49

By inspection of the spectrum, figure 8.16, the offset in ppm is ~310 ppm, thus the observation frequency is approximately $4000 = 310 \times \nu_0/10^6$ giving $\nu_0 = 12.9$ MHz – the actual spectrometer operating frequency was 12.76 MHz.

- b. At high enough spinning rates a resonance at the isotropic chemical shift is observed. Here, 4 kHz is fast enough to estimate of the isotropic chemical shift, $\delta_{so} \approx 10150$ ppm, directly from the spectrum. At lower spinning rates computer fitting of the static lineshape or spinning sidebands is used to obtain the principal components of the chemical shift tensor, δ_{11} , δ_{22} , δ_{33} . Here, estimates of δ_{11} and δ_{33} can be obtained by visual inspection of the spectrum at 1 kHz. Since a reasonable approximation of δ_{so} can be obtained from the 4 kHz spectrum, δ_{22} can be approximated from the definition of $\delta_{so} = \frac{1}{3}(\delta_{11} + \delta_{22} + \delta_{33})$. Finally, note that various conventions are used to order δ_{11} , δ_{22} , δ_{33} . Here $\delta_{11} \ge \delta_{22}$ $\ge \delta_{33}$ has been used giving $\delta_{11} \approx 10650$, $\delta_{22} \approx 10100$, $\delta_{33} \approx 9700$ ppm. The values determined by fitting the sidebands at a spinning rate of 1 kHz are: $\delta_{11} = 10551$, $\delta_{22} = 10048$, $\delta_{33} = 9793$ ppm.
- c. Cross-polarization builds up during the contact time when the irradiation used to transfer polarization between spins is "switched on" increasing the *S* spin signal, however, relaxation of the *S* spins also occurs during this time reducing the signal; signal intensity thus initially builds up due to CP and then falls away due to relaxation.



Fig. 8.16 ¹⁰³Rh{¹H} CP/MAS NMR spectrum of and CP build up curve for the symmetrical dimeric Rh cation [(H₂O)₄Rh(μ^2 -OH)₂Rh(H₂O)₄]. Adapted with permission from B. L. Phillips *et al., J. Amer. Chem. Soc.*, 2006, **128**, 3912-3913. Copyright © 2017, American Chemical Society.

Q7.2 Figure 7.24 shows the ¹³C{¹H} and ³¹P{¹H} spectra of [Pd(dppp)(dtc)]BF₄ (dppp = diphenylphosphinopropane, dtc = N,N-diethyldithiocarbamate) in solution and the solid state. Suggest an explanation for the differences between the solution and solid state spectra.

Answer

a. In solution molecular motions are fast but in the solid state are restricted. Thus, molecular tumbling in solution averages out dipolar interactions and chemical shift anisotropy resulting in sharp NMR lines. In the solid state spectra, however, the MAS spinning rate is not sufficient to average out these line broadening interactions resulting in broader spectral lines. Furthermore, in solution, vibrational motions of the ligands create a plane of symmetry through the nitrogen, palladium and the central carbon of the diphosphine ligand backbone that equivalences the two sides of the molecule. Thus, in solution a single chemical shift for phosphorus is observed and the ¹³C {¹H} spectrum shows a one shift for each of the types of carbon present (thiocarbamate methyl and methylene; *ortho, meta, para* and *ipso* carbons of the phenyl rings, C(1) and C(3) of the ligand backbone, and finally the central carbon.

Anchoring the Pd complex to a silica surface restricts these molecular motions severely, breaking the molecular symmetry; the two sides of the molecule become distinct and two resonances are observed ³¹P{¹H} CPMAS spectrum.

Similarly, additional resonances are seen in the solid-state ¹³C{¹H} NMR spectra as the previously equivalent aromatic carbons become distinct. Finally, as expected, the ¹³C resonance of the chloroform solvent is absent from the solid-state spectrum.

- b. Crystal packing can also result in inequivalencing of sites.
- **Q 7.3** In a cross polarization experiment a 9.4 T spectrometer was used; ¹H was the polarization source and ¹⁰⁹Ag was the polarization receiver.
- (a)What is the maximum signal enhancement that can be obtained neglecting relaxation effects?

Signal enhancement = $\gamma_I/\gamma_S = \gamma_{1H}/\gamma_{109Ag} = |26.75x10^7/-1.25x10^7| = 21.4$. (Or 20.4 if the negative γ correction applies as in INEPT)

- (b) The proton B_1 field used was 1.4 mT. What B_1 field is required at ¹⁰⁹Ag to fulfil Hartmann-Hahn matching condition at a MAS rate of 16 kHz?
- The Hartmann-Hahn matching condition is $B_1^{H} \gamma_{1H} = B_1^{Ag}\gamma_{109Ag} \pm nx2\pi x$ (MAS rate). The gyromagnetic ratios in Table 1.2 are given in $10^7 \text{rad}.\text{T}^{-1}.\text{s}^{-1}$ so ¹H will precess about the proton spin-lock field at $B_1^{H}x26.75x10^7/2\pi = 59.6$ kHz. n can take any positive or negative integer value, taking n = 1, the ¹⁰⁹Ag must precess about the ¹⁰⁹Ag spin-lock

field at 59.6 – 16 = 43.6 kHz whence $B_1^{Ag}g_{109Ag}$ = 43.6 kHz giving B_1^{Ag} = 43600x2 π/γ_{109Ag} = 21.9 mT.

Q7.4 The cathode (positive electrode) of a rechargeable battery is the electrode where reduction takes place during the discharge cycle; for lithium-ion cells this is the lithium-based electrode. Li-ion batteries typically comprise a lithium-doped cobalt oxide (LCO) positive electrode, and a hard carbon or graphite negative electrode, a porous separator, and a solution of a lithium salt in an organic liquid, figure 7.25. Overcharging of lithium ion batteries is reported to promote the growth of lithium metal dendrites on the cathode. Metal nuclei in metals are shielded by the metal conduction electrons so characteristically show large down field shifts (the Knight shift) whereas Li ions in solution typically have very small chemical shifts.

- Assign the ⁷Li solid-state NMR spectra shown in figure 7.25, the ⁷Li resonance of LCO is very broad and the electrolyte is 1 M LiPF₆.
- b. Is your assignment consistent with the formation of Li dendrites?

Answer

a. Spectrum (a) contains two resonances which can be assigned to the electrolyte (~ 0 ppm) and Li metal (a Knight shifted resonance at ~ 250 ppm). During the charge-discharge-recharge cycle (spectra b, c, and d respectively) a broad resonance around 40 ppm disappears then reappears and can be assigned to lithium carbides based on this behaviour and the absence of a Knight shift.

Overcharging results in a broad, Knight shifted resonance around 270 ppm that is assigned to dendritic lithium on the cathode. Finally, a very broad resonance around 0 ppm is attributed to lithium in LCO.

b. The appearance of a Knight shifted resonance on overcharging is consistent with overcharging causing dendritic lithium being deposited on the cathode.